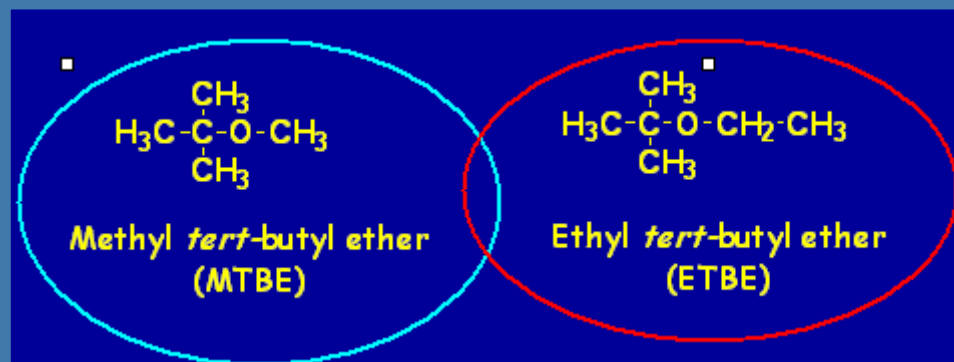


New tools of detection for the management of groundwater polluted by ether-fuels

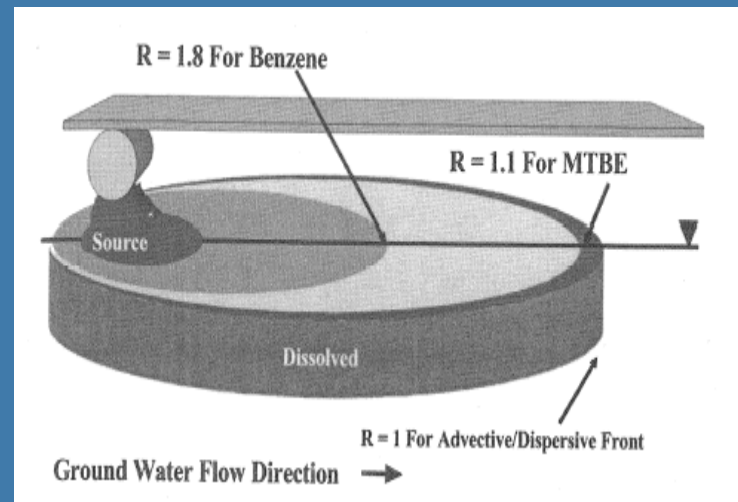


Françoise FAYOLLE-GUICHARD



Problems encountered with the use of MTBE/ETBE-supplemented gasoline

- Extent of MTBE (ETBE) plumes when an aquifer is polluted by MTBE/ETBE-supplemented gasoline (Wilson, 2003) :



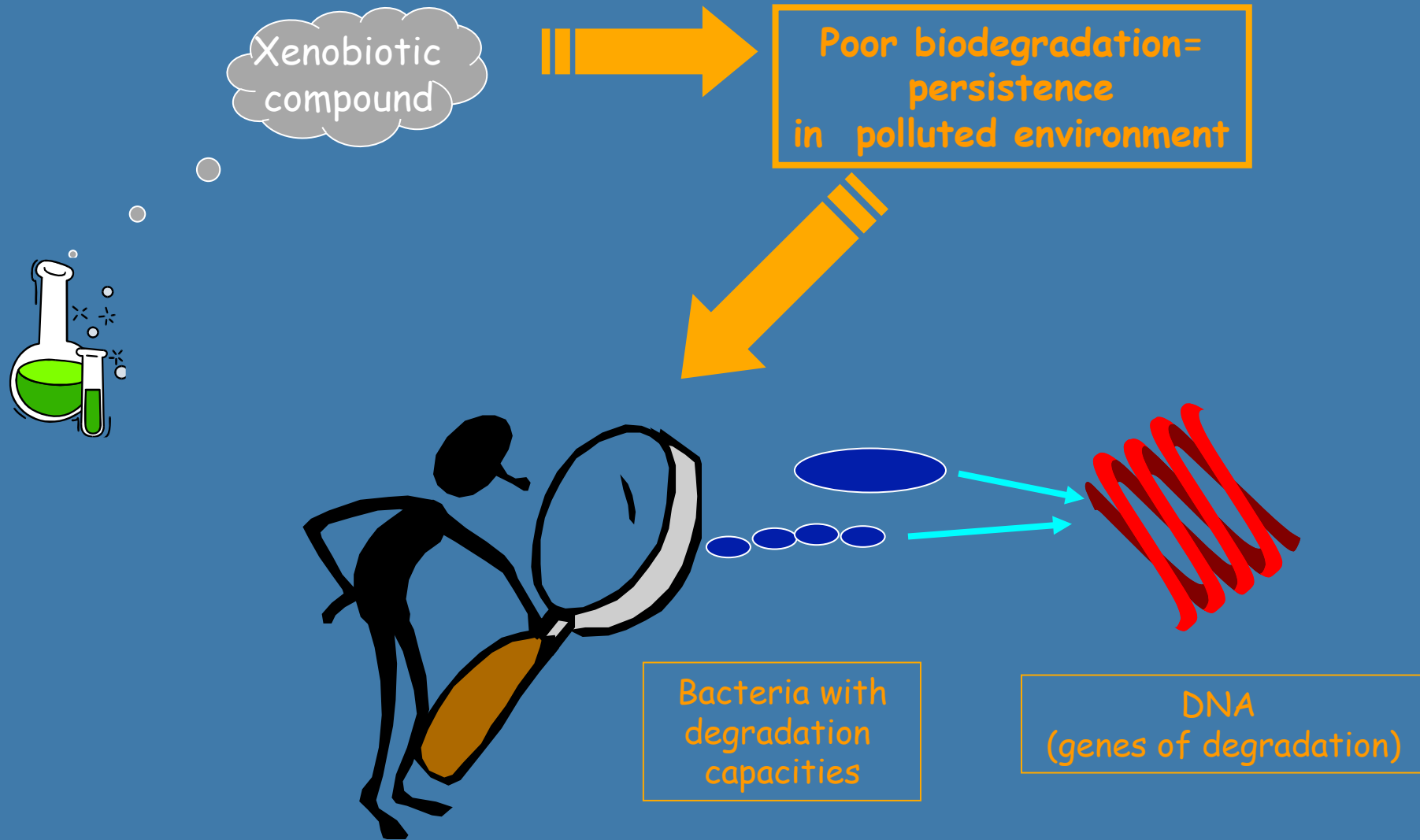
- High solubility (40 g/L for MTBE and 10 g/L for ETBE)
- Low adsorption on organic matter
- Recalcitrance to biodegradation



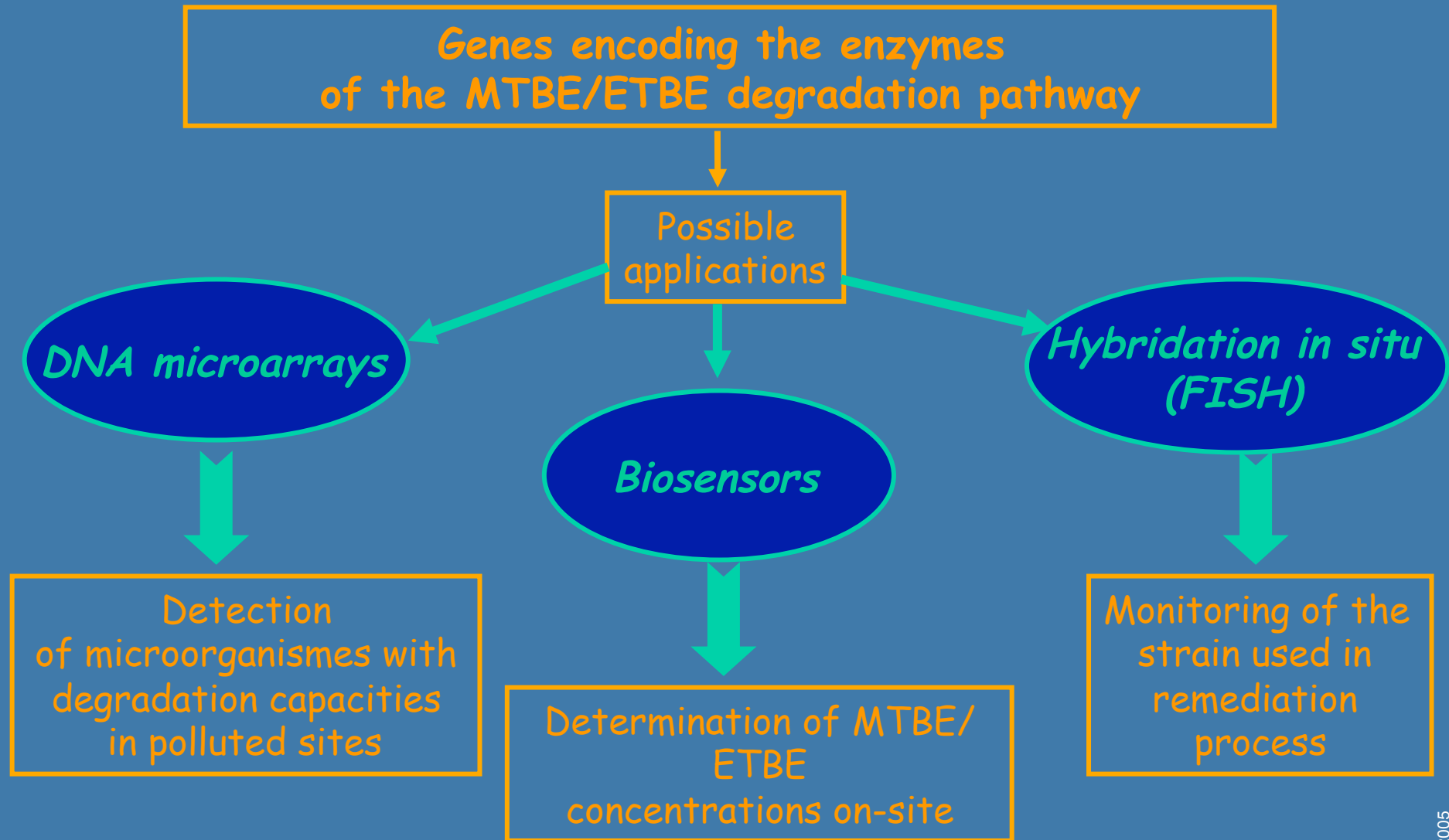
Questions posed to people involved in depollution of such sites

- How to estimate the potential for natural attenuation (if any) to both minimize and potentialize the actions that have to be undertaken for depollution ?
- How to monitor efficiently and at lower costs the ether-fuels concentrations in such large polluted areas ?
- How to clean the site if no biodegradation capacities have been detected ?

Interest to study strains with MTBE/ETBE biodegradation potential



Possible applications: new tools for environmental management



First question:

How to estimate the potential for natural attenuation?

IFP with Biotechnology Research Institute, Montréal
(Environmental Microbiology Group,
head: Dr Charles W. GREER)



Usual tools to estimate the potential for natural attenuation

-Classical respirometry tests:

- * Production of CO_2
 - * O_2 consumption
- } Not very useful in such a case

-Monitoring MTBE or ETBE residual concentrations during degradation tests using microflora present in the contaminated site:

⇒ Capacity of biodegradation
present *in situ* ?

⇒ Estimation of the production of
degradation intermediates (TBF, TBA)

- } -requires to have the
technical tools for such
analyses
-time-consuming



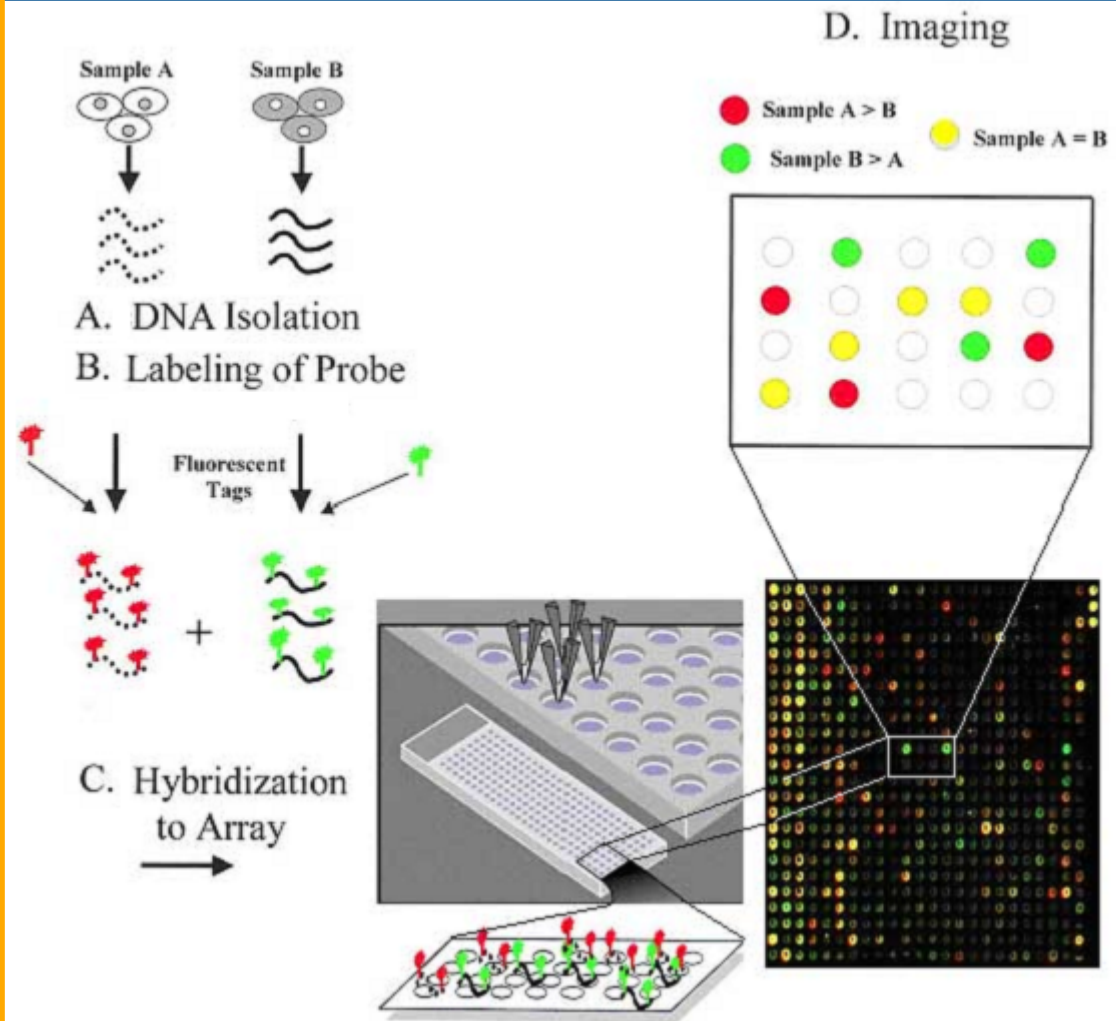
Biodegradability of ether-fuels by various microflora

Origin of microflorae	Biodegradation of MTBE (%)	Biodegradation of ETBE (%)	Biodegradation of TBA (%)	Biodegradation of TAME (%)
Activated sludge A	40	100	100	100
Activated sludge V	31	100	100	84
Activated sludge F	0	8	100	0
Soil M	37	100	100	0
Soil B	0	5	100	5
Soil G	4	0	100	0
Soil N	0	0	100	0

Principle of DNA microarrays

It is based on the following facts:

- DNA is a double-strand molecule,
- DNA sequences on each strand are complementary,
- the strands can easily be separated (DNA denaturation),
- each strand can then hybridized with a DNA sequence which is complementary,



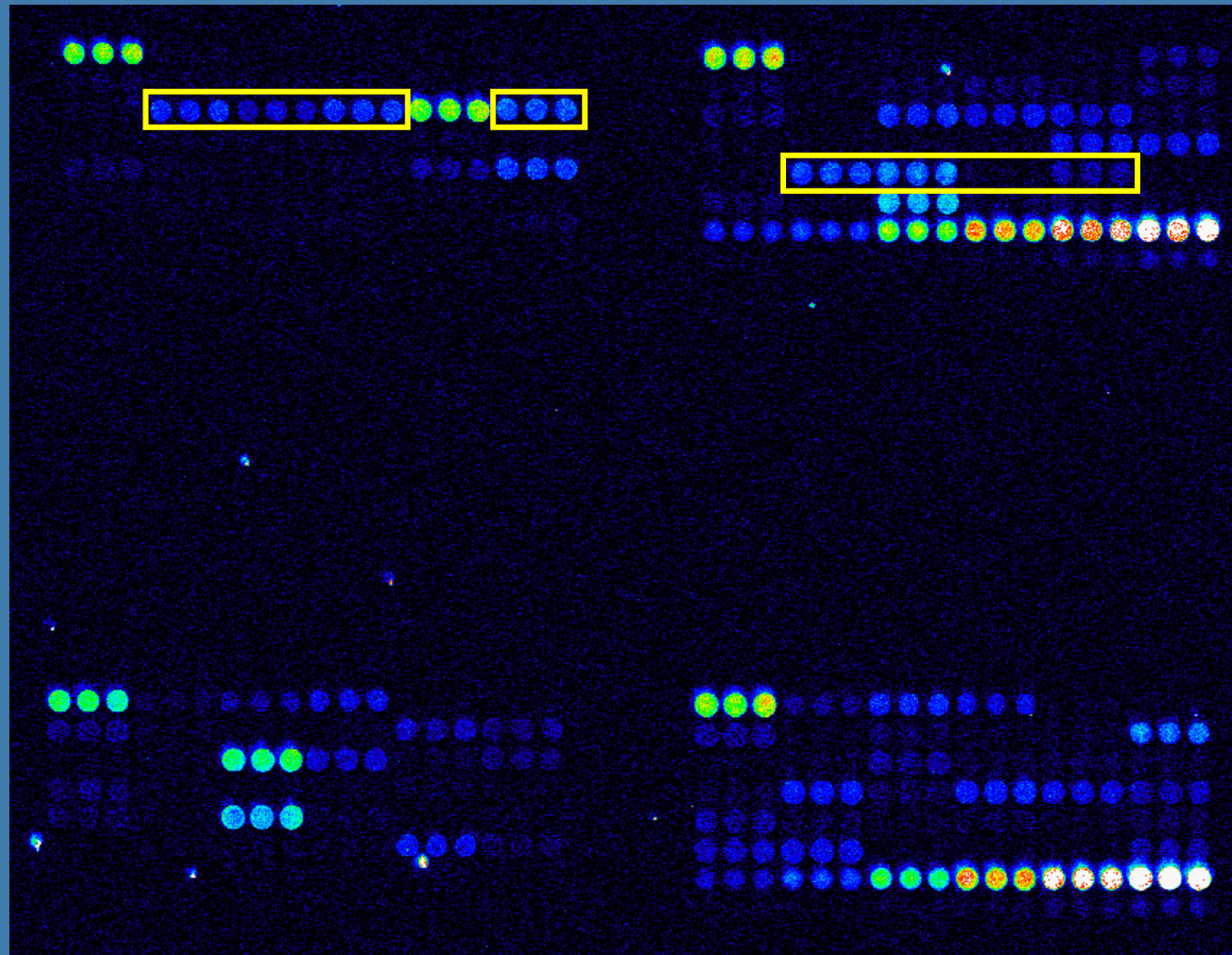


DNA microarrays: detection of alkane degradation genes

Biotechnology
Research Institute
(BRI), Canada
Group of
Charles GREER

Alkanes
degradation
genes

Soil DNA Extract Hybridized on a Catabolic Microarray :
Soil contaminated with petroleum hydrocarbons



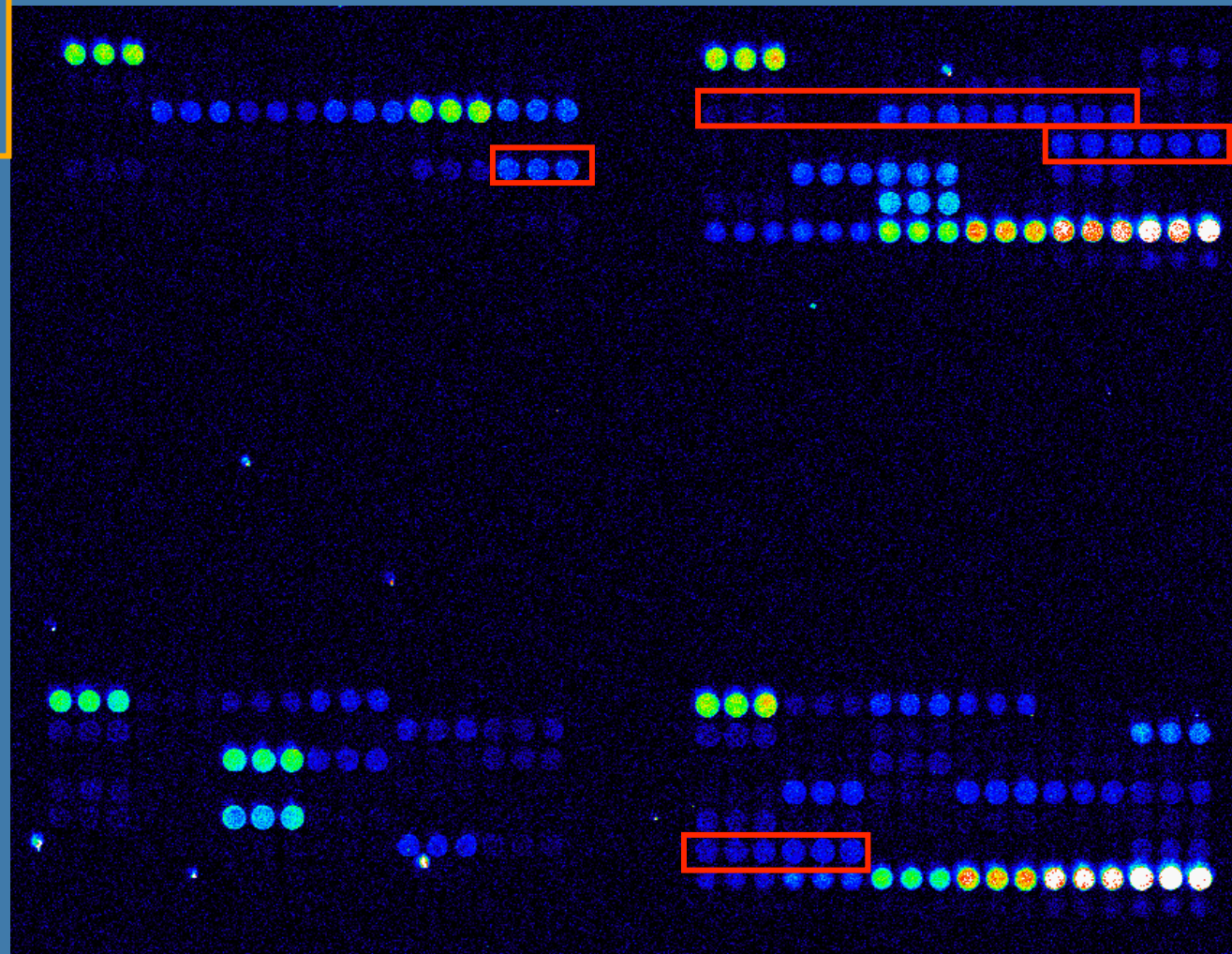


DNA microarrays: detection of naphthalene degradation genes

Biotechnology
Research Institute
(BRI), Canada
Group of
Charles GREER

Soil DNA Extract Hybridized on a Catabolic Microarray :
Soil contaminated with petroleum hydrocarbons

Alkanes
degradation
genes



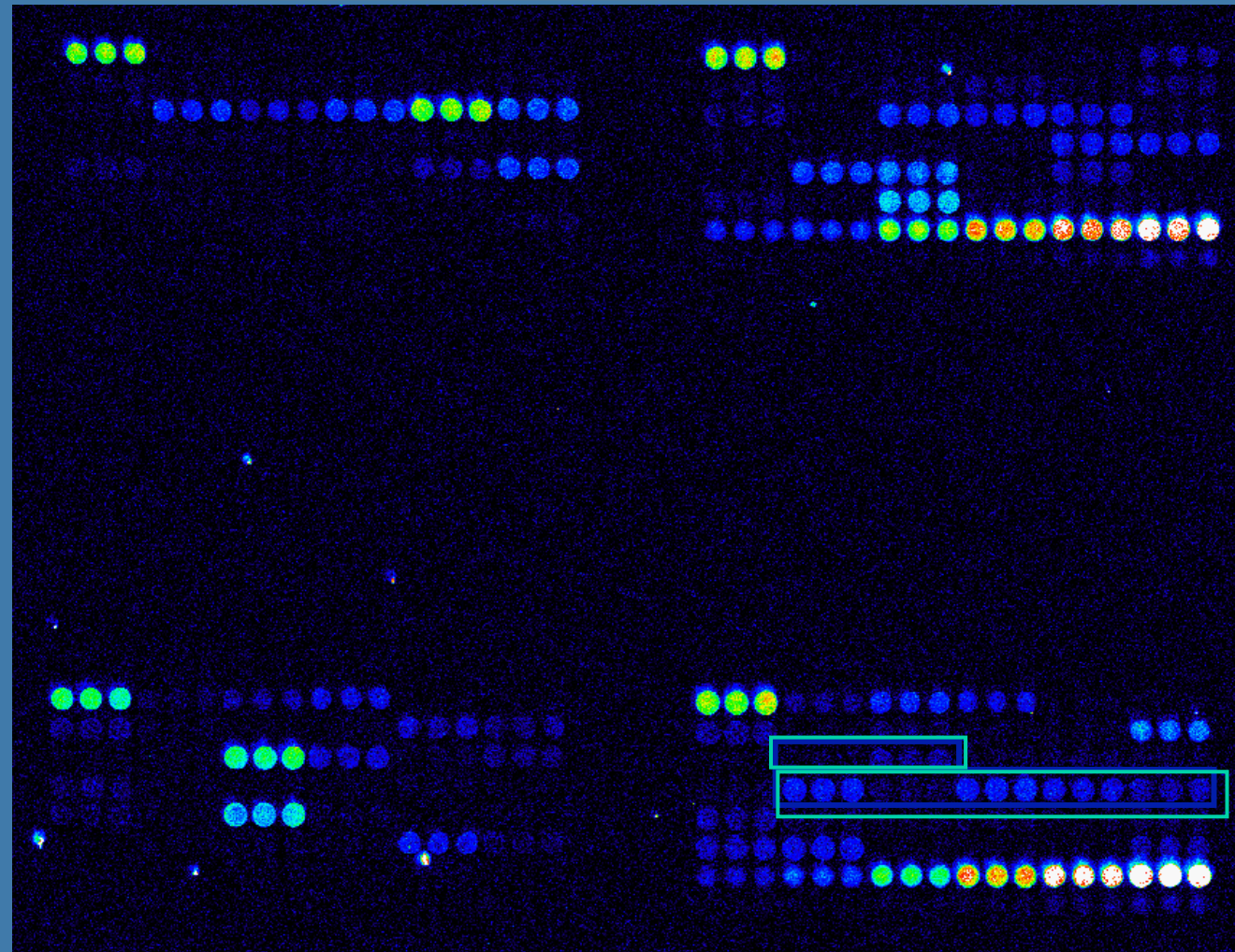


DNA microarrays: detection of monoaromatic degradation genes

Biotechnology
Research Institute
(BRI), Canada
Group of
Charles GREER

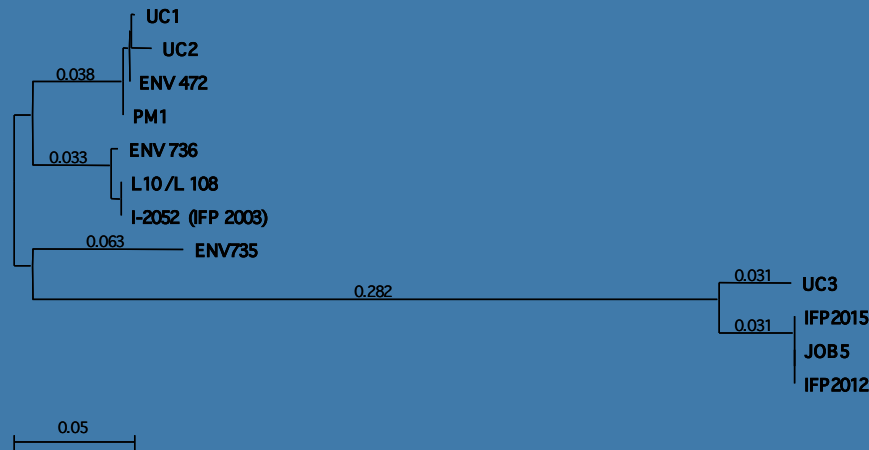
Monoaromatic
compounds
degradation
genes

Soil DNA Extract Hybridized on a Catabolic Microarray :
Soil contaminated with petroleum hydrocarbons



Ecology of MTBE biodegradation

- Isolation of strains with degradation capacities and their phylogenetic characterization:



Sequences of the 16S rDNA for detection of strains belonging to the different groups of the phylogenetic tree of MTBE-degraders

- Characterization of the enzymes involved in the degradation pathway
Isolation and cloning of the corresponding genes:

**eth* genes
**alk* genes
**mpd* genes



Microarrays for detection of MTBE/ ETBE degradation capacities



Several genes characterized



Validation step

-determination of limits for detecting the interesting genes
in samples containing complex DNA mixture.

-determination in parallel of:

- * the MTBE/ETBE/TBA degradation capacities on
samples from sites contaminated with ether fuels
- * the presence of the genes of interest in these
samples.

Second question:

How to monitor ether-fuels
concentrations in polluted sites ?

IFP with Center for Environmental Biotechnology,
Knoxville (USA)

(Head: Pr Gary SAYLER)

Usual tools to measure ether fuels concentrations

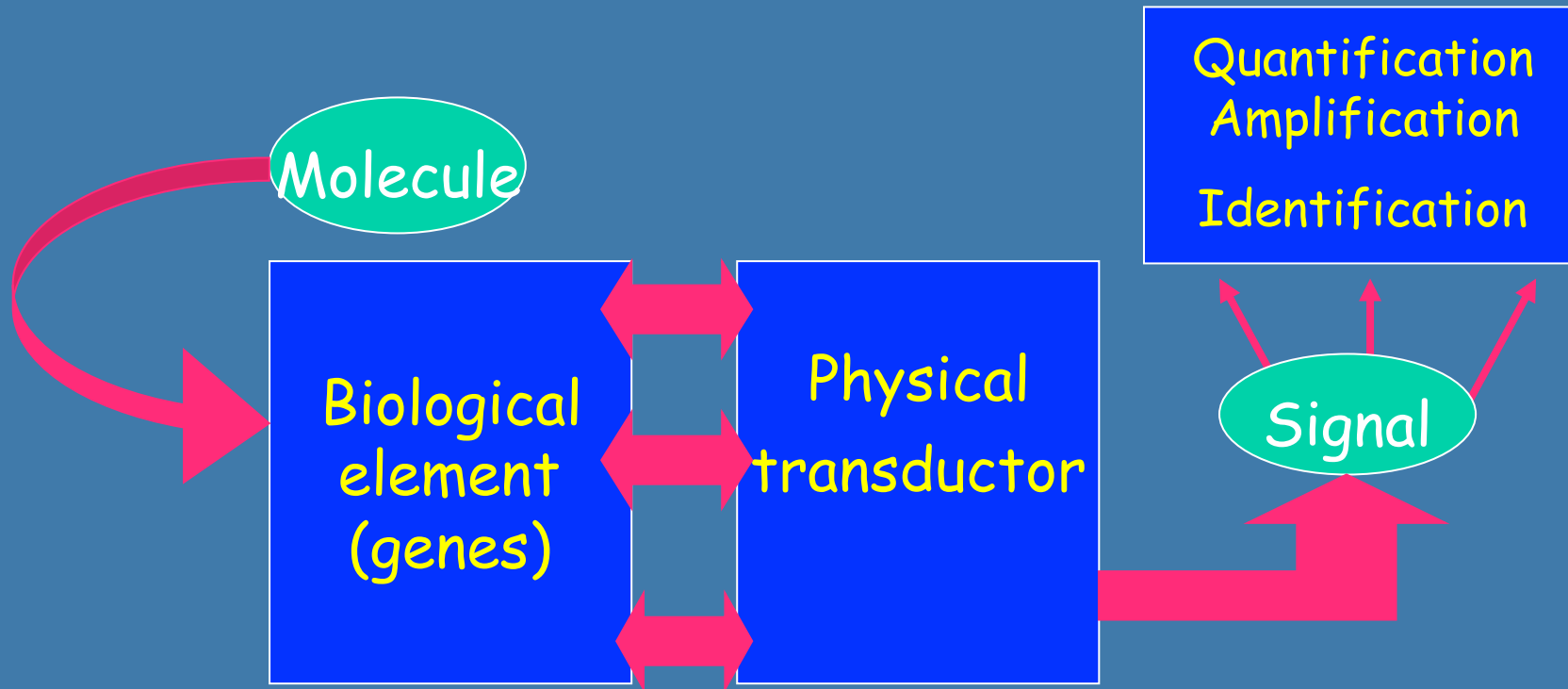
- Classical analytical method are very efficient (lower detection limit $0.02 \mu\text{g/L}$):
 - GC/MS
 - GC/FID
- Handling of numerous samples
- Analysis is costly and time-consuming

A new detection tool allowing specific determination and reliable quantification of ETBE or MTBE would be useful:



Bioluminescent biosensor

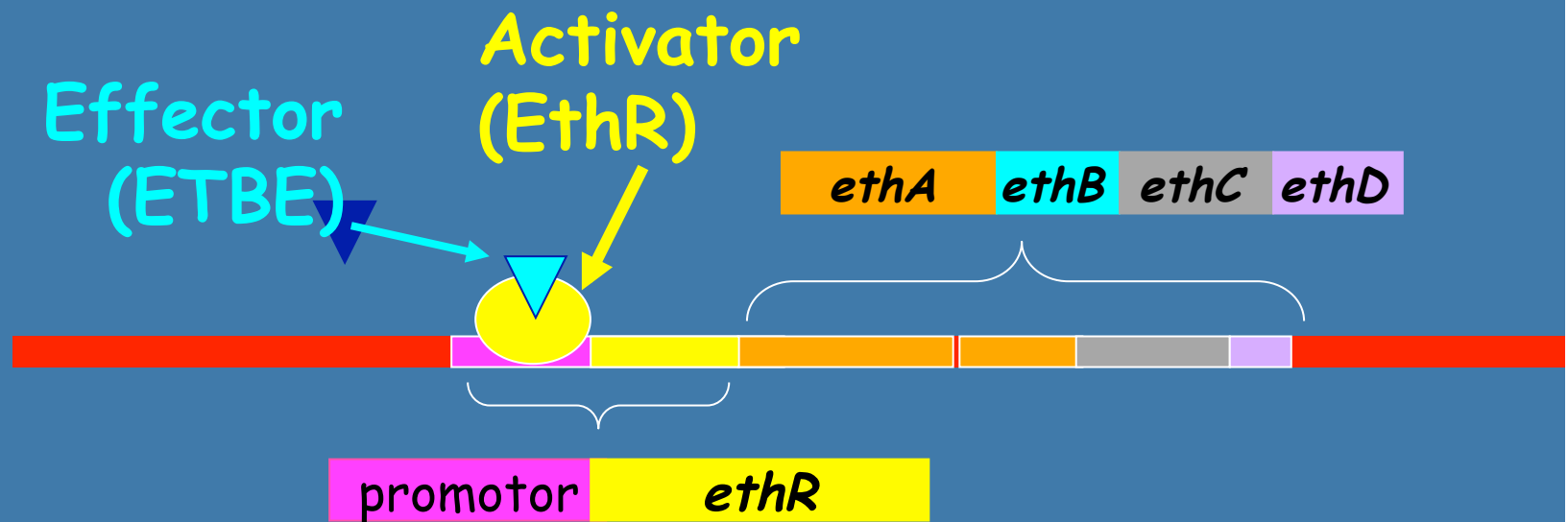
General principle of a biosensor



Elements required to build a bioluminescent biosensor

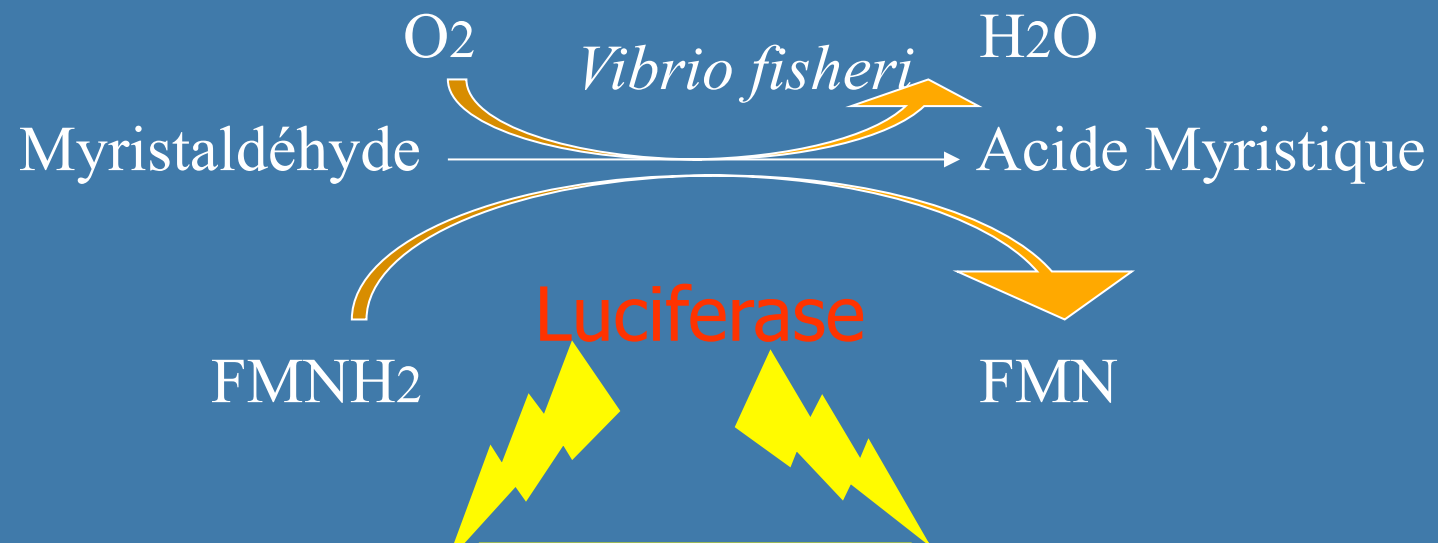
1

Genes for identification of ETBE:



Elements required to build a bioluminescent biosensor

2 Genes emitting light:



Genes of *V. fischeri* responsible for the synthesis of luciferase:

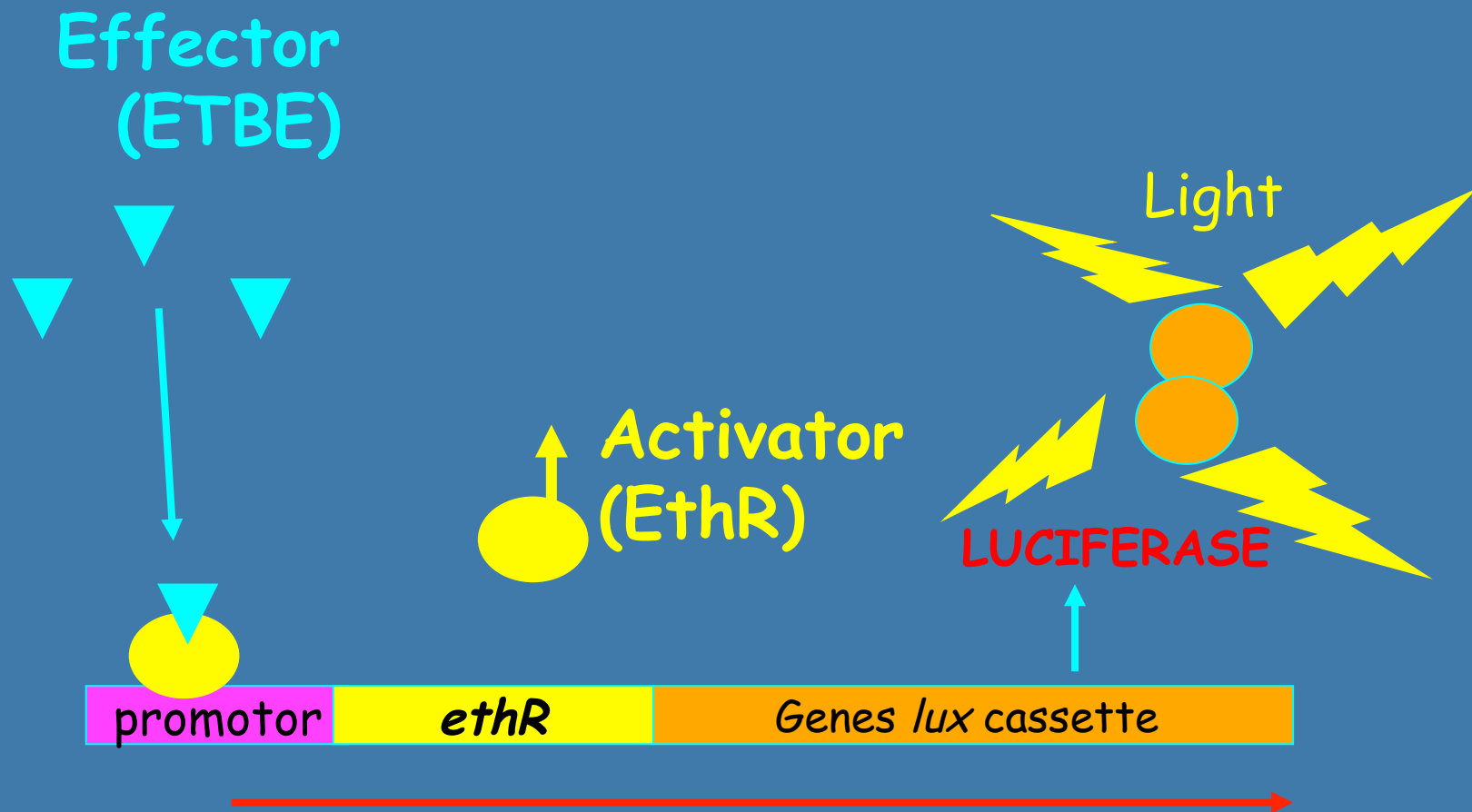
Cassette of
lux genes

= cassette *lux* commercially available

The bioluminescent biosensor to detect ETBE

3

Transcriptional fusion of genes:

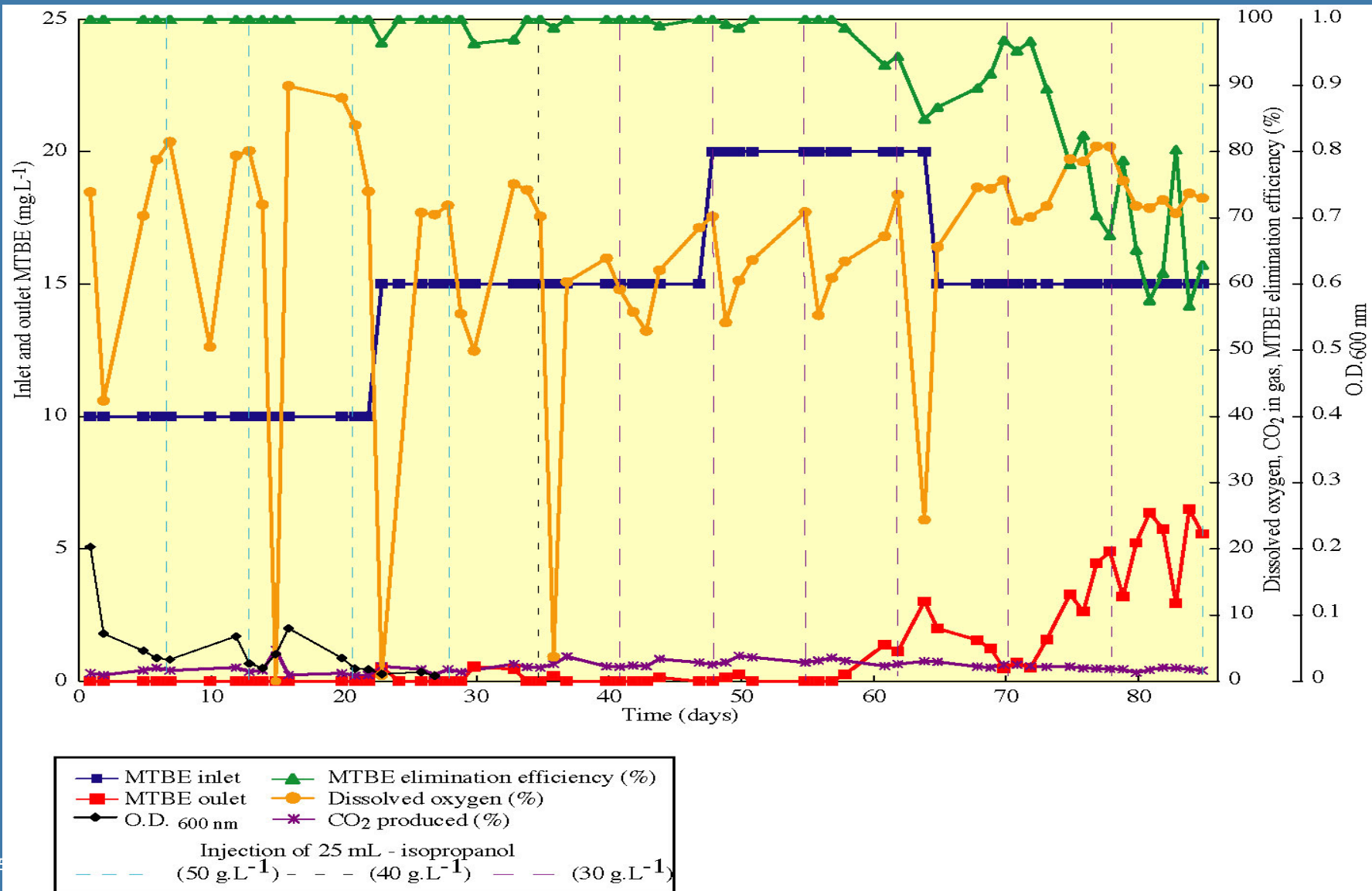


Third question:

How to clean a MTBE-contaminated site ?

IFP with Biotechnology Research Institute, Montréal
(Group Environment Biotechnology,
head: Dr Serge Guiot)

Bioremediation process using *M. austroafricanum* IFP 2012: biofilter

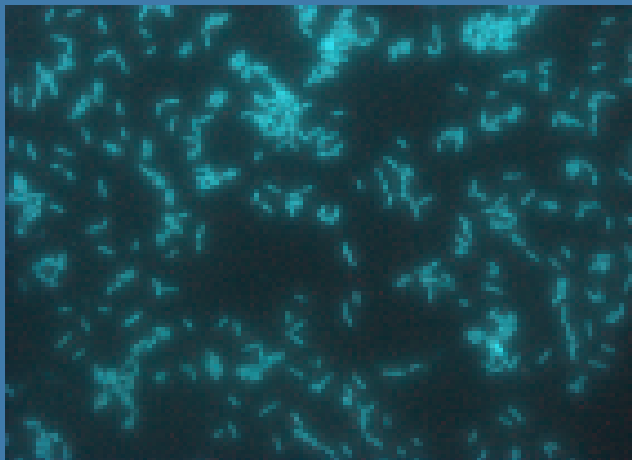




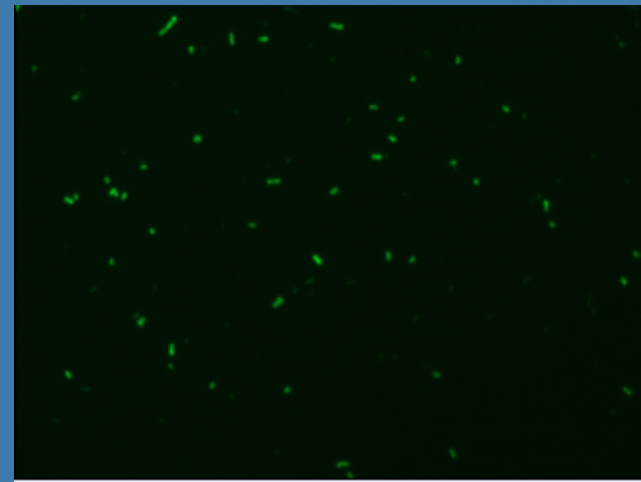
Monitoring *M. austroafricanum* IFP 2012 on the biofilter or in the site

Use of fluorescent in situ hybridization (FISH):

- labeling of probe (16S rDNA) with fluorescent tag
- specific for the detection of a given microorganism



DAPI



FISH



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Serge Guiot

-Center for Environmental Biotechnology, Knoxville, USA :

Steven Ripp, Gary Sayler.